

## General Flow Cytometry Protocol

Protocol may need local adjustments

### Direct Surface staining

1. Prepare the cells and adjust cell suspension to a concentration of  $1 - 5 \times 10^6$  cells/ml in cell staining buffer (eg PBS with  $\text{NaN}_3$  and BSA).
2. Ensure viability to be around 95%, not less than 90%.
3. Aliquot 100  $\mu\text{l}$  of cell suspension into as many test tubes as required.
4. Add the fluorochrome conjugated antibody (see the specific datasheets for recommended dilution).
5. Mix well and incubate for 15-30 min at room temperature or 4°C. This step will require optimization and must be done in the dark.
6. Wash the cells 1-3 times with staining buffer and centrifuge each time at 400 g for 5 minutes and discard the resulting supernatant.
7. Resuspend in remaining buffer and adjust volume with staining buffer.
8. Keep the cells in the dark on ice or at 4°C until acquisition.